Importance of Brevetoxin Determination

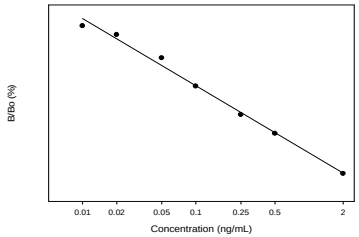
Neurotoxic shellfish poisoning (NSP) is caused by polyether toxins known as Brevetoxins. Brevetoxins (PbTxs) are produced by the dinoflagellate *Karenia brevis*, which causes harmful algal blooms (HABs) know as red tides. The Brevetoxins are toxic to fish, marine mammals, birds and humans, but not to shellfish. Contamination of shellfish with Brevetoxin has been associated with the presence of harmful algal blooms in various parts of the world.

Mortality events attributed to HABs have been documented for fish, manatee, dolphins, and seabirds. In man, NSP causes dose-dependent symptoms of diarrhea, nausea, and vomiting, chills, sweats, reversal of temperature, hypotension, arrhythmias, numbness, tingling, bronchoconstriction, paralysis, seizures, and coma.

The Brevetoxin ELISA allows the determination of 40 samples in duplicate determination. Only a few milliliters of sample are required. The test can be performed in less than 2 hours.

Performance Data

Test sensitivity: The limit of detection for Brevetoxin is calculated as: Xn +/- 3SD (n=20) and is equal to 0.05 ng/ml in water and 22.5 ng/gm in diluted shellfish (when using a dilution factor of 450). The concentration of residue necessary to cause 50% inhibition (50% B/B_0) is approximately 0.16 ng/mL. Determinations closer to the middle of the calibration range of the test yield the most accurate results.



Test reproducibility: Coefficients of variation (CVs) for standards: <10%, CVs for samples: <15%.

Selectivity: This ELISA recognizes Brevetoxin and other NSP toxins to varying degrees:

Cross-reactivities:

PbTx-3	100%
Deoxy PbTx-2	133%
PbTx-5	127%
PbTx-2	102%
PbTx-9	83%
PbTx-6	13%
PbTx-1	5%

No cross-reactivity was shown with any of the following common PSP shellfish toxins: saxitoxin, neosaxitoxin, dc-STX, gonyautoxins-1/4, gonyautoxins-2/3, B-2; B-1; C-1/2 and domoic acid.

Samples: Salt Water and shellfish samples (after recommended dilution) were tested for matrix effects in the ELISA. No matrix effects were determined.

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ABRAXIS

Brevetoxin (NSP) ELISA, Microtiter Plate

Enzyme-Linked Immunosorbent Assay for the Determination of Brevetoxin (NSP) in Water and Contaminated Samples

Product No. 520026

1. General Description

The Brevetoxin ELISA is an immunoassay for the quantitative and sensitive detection of Brevetoxin. Brevetoxin is one of the toxins associated with neurotoxic shellfish poisoning (NSP). This test is suitable for the quantitative and/or qualitative detection of Brevetoxin in water samples as well as shellfish samples. For shellfish samples a sample preparation is required. If necessary, positive samples can be confirmed by HPLC, GC/MS, or other conventional methods.

2. Safety Instructions

The standard solutions in this test kit contain small amounts of Brevetoxin (PbTx-3). In addition, the substrate solution contains tetramethylbenzidine and the stop solution contains diluted sulfuric acid. Avoid contact of stopping solution with skin and mucous membranes. If these reagents come in contact with the skin, wash with water.

3. Storage and Stability

The Brevetoxin ELISA should to be stored in the refrigerator (4–8°C). The solutions must be allowed to reach room temperature (20-25°C) before use. Reagents may be used until the expiration date on the box.

4. Test Principle

The test is a direct competitive ELISA based on the recognition of Brevetoxin by specific antibodies. Brevetoxin, when present in a sample, and a Brevetoxin enzyme-conjugate compete for the binding sites of sheep anti-brevetoxin antibodies that have been immobilized in the wells of a microtiter plate. After a washing step and addition of the substrate solution, a color signal is produced. The intensity of the blue color is inversely proportional to the concentration of Brevetoxin present in the sample. The color reaction is stopped after a specified time and the color is evaluated using an ELISA reader. The concentrations of the samples are determined by interpolation using the standard curve constructed with each run.

5. Limitations of the Brevetoxin ELISA, Possible Test Interference

Numerous organic and inorganic compounds commonly found in samples have been tested and found not to interfere with this test. However, due to the high variability of compounds that might be found in samples, test interferences caused by matrix effects can not be completely excluded. Mistakes in handling the test can also cause errors. Possible sources for such errors can be:

Inadequate storage conditions of the test kit, incorrect pipetting sequence or inaccurate volumes of the reagents, too long or too short incubation times during the immune and/or substrate reaction, extreme temperatures during the test performance (lower than 10°C or higher than 30°C).

The Abraxis Brevetoxin ELISA kit provides screening results. As with any analytical technique (GC, HPLC, etc.) positive samples requiring some action should be confirmed by an alternative method.

Working Instructions

A. Materials Provided

1. Microtiter plate coated with sheep anti-Brevetoxin
2. Standards PbTx-3 (8): 0, 0.010, 0.025, 0.05, 0.1, 0.25, 0.5, 2.0 ng/mL
3. Brevetoxin-HRP Conjugate, 6 mL
4. Sample Diluent (1X), 2 X 30 mL. Use to dilute samples
5. Wash Solution (5X) Concentrate, 100 mL
6. Color Solution (TMB), 12 mL
7. Stop Solution, 2 X 6 mL
8. Sea Water Pre-treatment Solution, 25 mL

B. Test Preparation

Micro-pipetting equipment and pipette tips for pipetting the standards and the samples are necessary. We recommend using a multi-channel pipette or a stepping pipette for adding the conjugate, the substrate solution and the stop solution in order to equalize the incubations periods of the standard solutions and the samples on the entire microtiter plate. Please use only the reagents and standards from one package lot in one test, as they have been adjusted in combination.

1. Adjust the microtiter plate and the reagents to room temperature before use.
2. Remove the number of microtiter plate strips required from the foil bag. The remaining strips are stored in the foil bag and zip-locked closed. Store the remaining kit in the refrigerator (4-8°C).
3. The standard solutions, enzyme conjugate, substrate and stop solution are ready to use and do not require any further dilutions.
4. Dilute the Wash Buffer at a ratio of 1:5. If using the entire bottle (100 mL), add to 400 mL of deionized or distilled water.
5. The stop solution should be handled with care as it contains diluted H_2SO_4.

C. Assay Procedure

1. Add 50 µL of the **standard solutions or the samples** (water) or **sample extracts** (shellfish) into the wells of the test strips according to the working scheme given. We recommend using duplicates or triplicates.
2. Add 50 µL of **enzyme conjugate** solution to the individual wells successively using a multi-channel pipette or a stepping pipette. Cover the wells with parafilm or tape and mix the contents by moving the strip holder in a rapid circular motion on the benchtop for about 30 seconds. Be careful not to spill contents. Incubate for 60 minutes.
3. After incubation, remove the covering and vigorously shake the contents of these wells into a sink. Wash the strips three times using the 1X washing buffer solution. Use at least a volume of 250 µL of washing buffer for each well and each washing step. Remaining buffer in the wells should be removed by patting the plate dry on a stack of paper towels.
4. Add 100 µL of **substrate solution** to the wells. The strips are incubated for 30 min at room temperature. Protect the strips from direct sunlight.
5. Add 100 µL of **stop solution** to the wells in the same sequence as for the substrate solution.
6. Read the absorbance at 450 nm using a microplate ELISA photometer within 15 minutes after the addition of the stopping solution.

D. Evaluation

The evaluation of the ELISA can be performed using commercial ELISA evaluation programs (4-Parameter (preferred) or Logit/Log). For manual evaluation, calculate the mean absorbance value for each of the standards. Calculate the $\%B/B_0$ for each standard by dividing the mean absorbance value for each standard by the Zero Standard (Standard 0) mean absorbance. Construct a standard curve by plotting the $\%B/B_0$ for each standard on the vertical linear (y) axis versus the corresponding Brevetoxin concentration on the horizontal logarithmic (x) axis on graph paper. $\%B/B_0$ for samples will then yield levels in ppb of Brevetoxin by interpolation using the standard curve. Samples showing lower concentrations of Brevetoxin compared to standard 1 (0.01 ng/mL) are considered as negative. Samples showing a higher concentration than standard 7 (2.0 ng/mL) must be diluted further to obtain more accurate results.

E. Additional Materials (not delivered with the test kit)

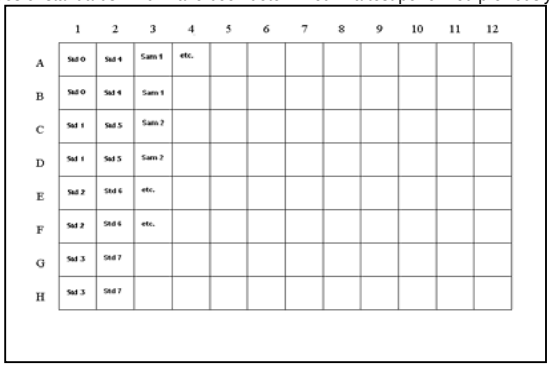
1. Micro-pipettes with disposable plastic tips (10-200, and 200-1000 µL)
2. Multi-channel pipette (10-250 µL) or stepper pipette with plastic tips (10-250 µL)
3. Microtiter plate washer (optional)
4. Microtiter plate reader (wave length 450 nm)
5. Shaker for microtiter plates (optional)

F. Working Scheme

The microtiter plate consists of 12 strips of 8 wells, which can be used individually for the test. The standards must be run with each test. Never use the values of standards which have been determined in a test performed previously.

Std 0-Std 7: Standards
0; 0.010; 0.025; 0.05; 0.1; 0.25
0.5; 2.0 (ng/mL) or ppb

Sam1, Sam2, etc.: Samples



G. Preparation of Samples

I. Mussels

1. Mussels are removed from their shells, washed with deionized water, thoroughly dried and homogenized (Waring blender, Polytron or equivalent).
2. A 1.0 gm portion of the homogenized mussels is then placed in a 40 mL glass vial.
3. Add 9.0 mL of a methanol/deionized water solution (9:1 v/v).
4. Vial is capped and hand shaken vigorously for 2 minutes.
5. Centrifuge mixture for 10 minutes at 3000 g. Collect the supernatant.
6. Remove 20 uL of collected extract and dilute to 1.0 mL with Sample Diluent (equals a 1:50 dilution).
7. Analyze diluted extracts as samples (Assay Procedure step 1).

The Brevetoxin concentration contained in the samples is determined by multiplying the concentration of the diluted extract by a factor of 450. Highly contaminated samples outside the range of the curve should be diluted further and re-analyzed.

II. Sea Water

1. Collect 2 mL of sea water sample in a glass container.
2. To prevent loss of Brevetoxin to the glass surface, immediately add 0.5 mL of Sea Water Pretreatment Solution, mix by hand.
3. Analyze preserved sample as samples (Assay Procedure step 1)

The Brevetoxin concentration contained in the sea water sample is determined by multiplying the concentration of the diluted sample by a factor of 1.25. Highly contaminated samples outside the range of the curve should be diluted in Sample Diluent (PN 205226), and re-analyzed. Additional Sample Diluent or Sea Water-Pretreatment Solution (PN 205227) can be purchased from Abraxis.