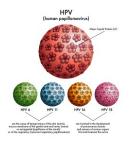
#### INTENDED USE

The **Rabbit anti- HPV18L1 IgG** ELISA Kit is an immunoassay suitable for detecting and quantifying IgG antibody activity specific for Human Papilloma Virus, subtype 18 L1 protein, in serum or plasma. This immunoassay is suitable for:

- Determining immune status relative to controls;
- Assessing efficacy of vaccines, including dosage, adjuvantcy, route of immunization and timing;

• Qualifying and standardizing vaccine batches & protocols. For research use only, not for diagnosis or therapeutic use.

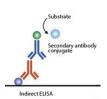
## **GENERAL INFORMATION**



Papillomavirus (HPV) causes cervical cancer, the third most common cancer in women worldwide. Lifetime incidence of HPV infection is estimated to be 80%. Like all papillomaviruses, HPVs establish productive infections only in keratinocytes of the skin or mucous membranes. Most infections become undetectable within 1–2 years and only a small fraction of infections with high-risk HPV fail

to clear, resulting in overt HPV persistence. Over 120 HPV types have been identified and are referred to by number; types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, and 59 are "high-risk" sexually transmitted HPVs. Neutralizing antibodies are expected to be the primary immune mechanism for protection against infection. Approved **vaccines** are now available to prevent infection by some HPV types: **Gardasil**, marketed by Merck, and **Cervarix**, marketed by GlaxoSmithKline. Both vaccines utilize recombinant L1 proteins and protect against initial infection with HPV types 16 and 18, which cause most of the HPV associated cancer cases. Newer vaccines, Gardasil9, has L1 proteins from 9 HPVs (6, 11, 16, 18, 31, 33, 45, 52, and 58). ADI has developed antibody ELISA kits for a single HPVL1 or various combinations of antigenic proteins to assess efficacy of vaccines.

## PRINCIPLE OF THE TEST



The Rabbit Anti-HPV18L1 IgG ELISA kit is based on the binding of anti-HPV18L1 IgG in samples to HPVL1 antigen immobilized on the microwells, and bound antibody is detected by anti-IgG-specific HRP-conjugate. After a washing step, TMB is added and color (blue) is developed, which is directly proportional to the amount of antibody

present in the sample. Stopping Solution is added to terminate the reaction, A450nm is measured using an ELISA reader. The presence of antibody in samples is determined relative to Calibrators.

#### PRODUCT SPECIFICATIONS

## Specificity

Purified recombinant (his tag; E.coli) HPV18L1 protein is used to coat the microwells; thus, no other antibody specificity is detectable in the assay. HPVL1s from HPV6, 11, 18, and 18 subtypes share ~50% sequence homology. The Anti-rabbit IgG HRP conjugate reacts specifically with rabbit IgG class antibodies; IgA, IgM and IgE antibody would not be measured above background signals.

## KIT CONTENTS

The microtiter well plate and all other reagents, if unopened, are

stable at 2-8<sup>O</sup>C until the expiration date printed on the box label. Stabilities of the working solutions are indicated under Reagent Preparation.

#### To Be Reconstituted: Store as indicated.

| Component   | Preparation Instructions   |
|---|--|
| Wash Solution<br>Concentrate (100x)<br>Cat. No. WB-100,<br>10ml                       | Dilute the entire volume 10ml + 990ml<br>with distilled or deionized water into a<br>clean stock bottle. Label as <b>Working</b><br><b>Wash Solution</b> and store at 4°C for<br>long term and ambient temp. for short<br>term.  |
| Sample Diluent<br>Concentrate (20x)<br>Cat. No. SD-20T,<br>10ml                       | Dilute the entire volume, 10ml + 190ml<br>with distilled or deionized water into a<br>clean stock bottle. Label as <b>Working</b><br><b>Sample Diluent</b> (WSD) and store at 2-<br>8°C until the kit lot expires or is used<br>up.  |
| Anti-Rabbit IgG-<br>HRP Conjugate<br>Concentrate (100x)<br>Part: H-RbG-211,<br>0.15ml | Peroxidase conjugated anti-rabbit IgG<br>in buffer with detergents and<br>antimicrobial as stabilizers. Dilute fresh<br>as needed; 10ul of concentrate to 1ml<br>of <b>Working Sample Diluent</b> is<br>sufficient for 1 8-well strip. Use within<br>the working day and discard. Return<br>100X to 2-8°C storage. |

### Ready For Use: Store as indicated on labels.

| Component                                 | Part                                      | Amt                                      | Contents  |
|---|---|--|---|
| HPV18<br>Antigen<br>Coated<br>Strip Plate | 550-<br>HPV18                             | 8-well<br>strips<br>(12)                 | Coated with recombinant<br>HPV18L1 protein, and<br>post-coated with<br>stabilizers.                     |
| Anti-HPV18 C                              | alibratore                                |  |   |
| 1 U/ml<br>2.5 U/ml<br>5 U/ml<br>10 U/ml   | 550222B<br>550222C<br>550222D<br>550222E  | 0.65 ml<br>0.65 ml<br>0.65 ml<br>0.65 ml | Four (4) vials, each<br>containing anti-<br>HPV18L1; in buffer<br>with antimicrobial as<br>stabilizers. |
| Low NSB<br>Sample<br>Diluent<br>(LNSB)    | TBTm<br><b>Not</b> for<br>HRP<br>dilution | 30 ml                                    | Buffer with protein,<br>detergents and anti-<br>microbial.<br>Use as is for sample<br>dilution          |
| TMB<br>Substrate                          | 80091                                     | 12 ml                                    | Chromogenic<br>substrate for HRP<br>containing TMB and<br>peroxide.                                     |
| Stop<br>Solution                          | 80101                                     | 12 ml                                    | Dilute sulfuric acid.   |

#### Materials Required But Not Provided:

- Pipettors and pipettes that deliver 100ul and 1-10ml.
- Disposable glass or plastic 5-15ml tubes for diluting samples and Anti-Rabbit IgG HRP Concentrate.
- Stock bottle to store diluted Wash Solution; 0.2 to 1L.
- Distilled or deionized water to dilute reagent concentrates. Microwell plate reader at 450 nm wavelength and ELISA plate washer

Page 2

## ASSAY DESIGN AND SET-UP

#### Sample Collection and Handling

Serum and other biological fluids may be used as samples with proper dilution to avoid solution matrix interference. For **serum**, collect blood by venipuncture, allow clotting, and separate the serum by centrifugation at room temperature. For other samples, clarify the sample by centrifugation and/or filtration prior to dilution in Sample Diluent. If samples will not be assayed immediately, store refrigerated for up to a few weeks, or frozen for long-term storage.

#### Antibody Stability & Dilution

Initial dilution of serum into **Working Sample Diluent** (WSD) is recommended to stabilize antibody activity. This enhances reproducible sampling, and stabilizes the antibody activity for years, stored refrigerated or frozen. Further dilution into **Low NSB Sample Diluent** (LNSD), which provides the lowest assay background, should be at least 10 times the initial dilution and performed the same day as the assay.

Example: Initial (1/5): **10**ul serum + **40**ul WSD [or 0.1ml + 0.4ml]

Further (1/50): **10**ul initial (1/5) + **90**ul LNSD (1/50)

#### Assay Design

Review Interpretation of Results and Limits of the Assay (p5-7) before proceeding:

- Select the proper sample dilutions accounting for expected potency of positives and minimizing non-specific binding (NSB) and other matrix effects; for example, net signal for non-immune samples should be lower than the 1 U/mI Calibrator. This is usually 1/100 or greater dilution for rabbit serum/plasma with normal levels of IgG and IgM.
- Run a Sample Diluent Blank. This signal is an indicator of proper assay performance, especially of washing efficacy, and is used for net OD calculations, if required. Blank OD should be <0.3.</li>
- Run a set of Calibrators, which validate that the assay was performed to specifications: 10 U/ml should give a high signal (>1.5 OD); 1 U/ml should give a low signal which can be used to discriminate at the Positive/Negative threshold (see Interpretation of Results, p. 5).

## Plate Set-up

Bring all reagents to room temperature (18-30° C) equilibration (at least 30 minutes).

- Determine the number of wells for the assay run. Duplicates are recommended, including 8 Calibrator wells and 2 wells for each sample control to be assayed.
- Remove the appropriate number of microwell strips from the pouch and return unused strips to the pouch. Reseal the pouch and store refrigerated.
- Add 200-300ul Working Wash Solution to each well and let stand for about 5 minutes. Aspirate or dump the liquid and pat dry on a paper towel before sample addition.

## Assay Procedure

ALL STEPS ARE PERFORMED AT ROOM TEMPERATURE. After each reagent addition, gently tap the plate to mix the well contents prior to beginning incubation.

- 1. 1<sup>st</sup> Incubation [100ul 60 min; 4 washes]
- Add 100ul of calibrators, samples and controls each to predetermined wells.
- Tap the plate gently to mix reagents and incubate for 60 minutes.
- Wash wells 4 times and pat dry on fresh paper towels. As an alternative, an automatic plate washer may be used. Improper washes may lead to falsely elevated signals and poor reproducibility.

[100ul - 30 min; 5 washes]

- o Add 100ul of diluted Anti-Rabbit IgG HRP to each well.
- Incubate for 30 minutes.

2<sup>nd</sup> Incubation

2.

3.

• Wash wells 5 times as in step 2.

#### Substrate Incubation [100ul – 15 min]

- Add 100ul TMB Substrate to each well. The liquid in the wells will begin to turn blue.
- Incubate for 15 minutes in the dark, e.g., place in a drawer or closet.

Note: If your microplate reader does not register optical density (OD) above 2.0, incubate for less time, or read OD at 405-410 nm (results are valid).

- 4. Stop Step [Stop: 100ul]
- Add 100ul of Stop Solution to each well.
- Tap gently to mix. The enzyme reaction will stop; liquid in the wells will turn yellow.

## 5. Absorbance Reading

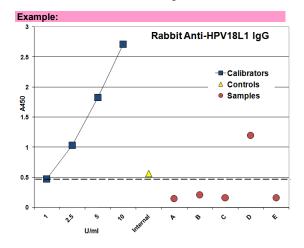
- Use any commercially available microplate reader capable of reading at 450nm wavelength. Use a program suitable for obtaining OD readings, and data calculations if available.
- Read absorbance of the entire plate at 450nm using a single wavelength within 30 minutes after Stop Solution addition. If available, program to subtract OD at 630nm to normalize well background.

## INTERPRETATION OF RESULTS

## Method A. Antibody Activity Threshold Index

Compare Samples to 1 U/ml Calibrator or Internal Control

#### = Positive/Negative Cut-off.



## Results

The **sensitivity** of the assay to detect anti-HPV18 IgG, from either natural infection or vaccination, is controlled so that the **1 U/mI Calibrator** represents a threshold OD for most true positives in rabbit serum diluted to 1:100 or greater. Visual inspection of the data in the above graph shows the following:

Calibrators – dilution curve of an anti-HPV antibody, derived from HPV18L1 vaccination, shows the OD range of the assay; high value indicates optimal sensitivity of the assay.
1 U/mI: a 'Cut-off' line has been drawn to indicate a threshold distinguishing between Positive/Negative. This is not a clear-cut threshold, rather a low OD area that could represent either low positives or high background negatives.

**Internal Control** – a low level positive from an immunized animal that represents the lab's experience in distinguishing low positive from negative samples. This should be run in each assay to supplement the 1 U/ml Calibrator for Positive/Negative discrimination purposes.

Samples A,B,C,D,E – 4 samples (A, B, C, E) are <u>negative</u>: below the threshold; 1 sample (D) is <u>positive</u>: clearly above the threshold.

The 1 U/ml Calibrator can be used to calculate a **Threshold Index** that numerically discriminates Positive/Negative:

Divide each Sample net OD by the 1 U/ml Calibrator net OD. Values above 1.0 are a measure of **Positive** Antibody Activity; below 1.0 are **Negative** for antibody.

## INTERPRETATION OF RESULTS (cont)

### Method B. Positive Index

Experimental sample values may be expressed relative to the values of Control or Non-immune samples, by calculation of a **Positive Index**. One typical method is as follows:

- Calculate the net OD mean + 2 SD of the Control/Nonimmune samples = Positive Index.
- Divide each sample net OD by the Positive Index. Values above 1.0 are a measure of **Positive** Antibody Activity; below 1.0 are **Negative** for antibody.

A sample value would be **Positive** if significantly above the value of the pre-immune serum sample or a suitably determined nonimmune panel or pool of samples, tested at the same sample dilution.

This calculation also **quantifies** the positive Antibody Activity level, assigning a higher value to samples with higher Antibody Activity, and vice versa.

|               |         | Assay<br>Net OD |         | Calculated<br>Antibody Activity |  |
|---------------|---------|-----------------|---------|---------------------------------|--|
| Sample        | Control | Exptl           | Control | Exptl                           |  |
| 1             | 0.244   | <b>C</b> 2.293  | 0.57    | 5.34                            |  |
| 2             | 0.204   | <b>C</b> 1.490  | 0.48    | 3.47                            |  |
| 3             | 0.237   | <b>C</b> 0.833  | 0.55    | 1.94                            |  |
| 4             | 0.26    | <b>C</b> 0.326  | 0.61    | 0.76                            |  |
| 5             | 0.388   | <b>P</b> 1.106  | 0.90    | 2.58                            |  |
| 6             | 0.407   | I 0.310         | 0.95    | 0.72                            |  |
| 7             | 0.288   | <b>E</b> 0.672  | 0.67    | 1.56                            |  |
| 8             | 0.263   | E 0.363         | 0.61    | 0.85                            |  |
| 9             | 0.322   | <b>E</b> 0.560  | 0.75    | 1.31                            |  |
| 10            | 0.343   | <b>E</b> 0.490  | 0.80    | 1.14                            |  |
| Mean          | 0.295   |                 |         |                                 |  |
| SD            | 0.067   |                 |         |                                 |  |
| Mean<br>+2 SD | 0.429   | = Positive      | Index   |                                 |  |

## <u>Results</u>

Evemple

 $\begin{array}{l} \textbf{Experimental Samples are represented as follows:}\\ \textbf{C} - Calibrator\\ \textbf{P} - Positive Control\\ \textbf{I} - Internal Control; lab's threshold positive serum\\ \textbf{E} - Experimental sample \end{array}$ 

## Assay Sensitivity

The HPV18 antigen coating level, Low NSB Sample Diluent, and HRP conjugate concentration are optimized to differentiate anti-HPV18 IgG from background (non-antibody) signal with rabbit serum or plasma samples diluted 1:100.

## ASSAY PERFORMANCE

#### Method C. Titers from Sample Dilution Curves

The titer of antibody activity calculated from a dilution curve of each sample is recommended as the most accurate quantitative method. Best precision can be obtained using the following guidelines:

- Use an OD value Index in the mid-range of the assay (2.0 – 0.5 OD); this provides the best sensitivity and reproducibility for comparing experimental groups and replicates. An arbitrary 1.0 OD is commonly used.
- Prepare serial dilutions of each sample to provide a series that will produce signals higher and lower than the selected index. With accurate diluting, duplicates may not be required if at least 4 dilutions are run per sample.
- A 5-fold dilution scheme is useful to efficiently cover a wide range which produces ODs both above and below 1.0 OD. The dilution scheme can be tightened to 3-fold or 2-fold for more precise comparative data.
- 4. The Positive and Sensitivity Control values can be used to normalize inter-assay values.

#### Calculations

- On a log scale of inverse of Sample Dilution as the x-axis, plot the OD values of the two dilutions of each positive sample having ODs above and below the OD value of the Index (arbitrary or selected Calibrator).
- From a point-to-point line drawn between the two sample ODs, read the dilution value (x-axis) corresponding to the OD of the selected Index
   = IgG Antibody Activity Units

#### Limits of the Assay

- The assay detects and quantifies IgG antibodies directed to the L1 protein. Animals may be exposed to the virus without producing antibodies specific to L1.
- Anti-HPV16 antibody levels of an immunized animal may be below detection threshold related to the time course of the infection, e.g., too early for positive titer development.
- The sensitivity of the assay may be increased to perhaps convert a borderline sample to a positive by using a lower dilution of the sample, e.g., 1/50. The values of negatives may increase, so an alternative threshold should be established using known negatives to develop a **Positive Index** (page 6), or by using known **Internal Controls** as discriminator for a **Threshold Control** (instead of the kit 1 U/ml Calibrator Control)

## PRECAUTIONS AND SAFETY INSTRUCTIONS

Calibrators, Sample Diluent, and Antibody HRP contain bromonitrodioxane (BND: 0.05%, w/v). Stop Solution contains dilute sulfuric acid. Follow good laboratory practices, and avoid ingestion or contact of any reagent with skin, eyes or mucous membranes. All reagents may be disposed of down a drain with copious amounts of water. MSDS for TMB, sulfuric acid and BND can be requested or obtained from the ADI website: Sample Diluent and anti-Protein G-HRP contain Proclin 300 (0.05%, v/v).

Instruction Manual No. M-550-218-PRG

# Rabbit Anti-HPV18L1 IgG ELISA Kit

## Cat. # 550-218-PRG, 96 tests

For the Detection and Quantitation of Anti-HPV18L1 IgG in Rabbit Serum/Plasma or other biological fluids

For research use only, not for diagnostic or therapeutic use.



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